

The effect of aerobic training with Citrus aurantium L. on Sirtuin 1 and peroxisome proliferator-activated receptor gamma coactivator 1-alpha gene expression levels in the liver tissue of elderly rats

Zinab Shykholeslami¹, Ahmad Abdi^{1*}, Alireza Barari¹, Seyed Ali Hosseini²

1. Department of Sport Physiology, Ayatollah Amoli Branch, Islamic Azad University, Amol, Iran.

2. Department of Sport Physiology, Marvdasht Branch, Islamic Azad University, Marvdasht, Iran.

Article Type: Original Article

Article History:

Received: 10 Jun 2019 Revised: 29 Jul 2019 Accepted: 29 Oct 2019

*Correspondence:

Ahmad Abdi, Department of Sport Physiology, Ayatollah Amoli Branch, Islamic Azad University, Amol, Iran a.abdi58@gmail.com



Abstract

Background and objective: Proper nutrition and exercise are two effective factors in improving liver function in old age. The aim of this study was to investigate the effect of aerobic training (T) with consumption of Citrus aurantium (CA) on Sirtuin 1 (SIR1) and peroxisome proliferator-activated receptor gamma coactivator 1-alpha (PGC-1 α) gene expression levels in the liver tissue of elderly rats.

Methods: In this experimental study, 25 elderly female rats were with mean weight of 231.12±22.43 g placed in five groups of 5 rats, including 1) control, 2) sham, 3) CA, 4) T and 5) T+CA. Over the course of eight weeks, groups 4 and 5 ran on the treadmill three sessions per week at 65 to 75 percent of the maximum running speed, and groups 3 and 5 received 300 mg/kg/day of CA extract peritoneally. Forty- eight hours after the last training session and CA consumption, SIR1 and PGC-1 α gene expression levels in the liver tissue were measured by real-time PCR method.

Results: T had a significant effect on increasing SIR1 (P = 0.009) and PGC-1 α (P = 0.001) gene expression levels; CA had a significant effect on reducing SIR1 gene expression levels and increasing PGC-1 α (P = 0.001); T + CA had a greater effect on increasing PGC-1 α gene expression levels than T and CA (P = 0.001).

Conclusion: Although eight weeks of T and CA consumption alone appear to improve PGC-1 α gene expression levels in the liver tissue of elderly rats, concurrent T and CA consumption has more favorable effects than each of them alone.

Keywords: Training, Citrus aurantium, Liver, Metabolism, Elderly

Copyright© 2018, **Jorjani Biomedicine Journal** has published this work as an open access article under the terms of the Creative Commons Attribution License (<u>http://creativecommons.org/licenses/by-nc/4.0/</u>) which permits non-commercial uses of the work while it is properly cited.



Introduction

Aging is a decrease in body readiness as a result of progressive decline in function and the ability to maintain homeostasis. The ability of tissues to repair and maintain their natural structure and function decreases with age. Therefore, older people are more prone to chronic liver disease (1). The liver is an organ that has a very vital function, however liver function decreases with age (2). The liver regulates energy metabolism through homeostasis, hepatic glucose and fat biosynthesis and steroid destruction, and glucose signaling. Therefore, the liver plays an important role in nutritional interventions in aging and age-related diseases. On the other hand, impaired hepatic metabolism brings about insulin resistance, diabetes mellitus, and non-alcoholic fatty liver disease (1). While the liver is highly resistant to aging, the available evidence suggests that the liver experiences all the symptoms of cellular aging. The aging process in the liver leads to impaired mitochondrial function and dietsensitive pathways, leading to cellular aging and inflammation. These events lead to several changes in the phenotype of the liver and impair the function of liver cells (3). Aging liver cells increases in the accumulation of fat droplets, reduces the oxidation capacity of mitochondria, and increases the reactive species of oxygen (4). One of the most important changes is the change in FOXO, which is a key factor in aging (5). FOXO is regulated bv NAD⁺/SIRT1, insulin-like growth factor 1, AMPK, and oxidative stress, all of which are affected by aging. SIRT1 is involved in the regulation of several key proteins related to aging, glucose homeostasis, lipid metabolism, autophagy, inflammation, apoptosis, and cellular changes (6, 7). SIRT1 activity is

dependent on NAD⁺ and has been shown to play a role in the beneficial effects of CR on longevity (8). Age-related decline in NAD⁺ impairs SIRT1 function (7). Inactive activity can accelerate the aging process by lowering certain hormones and increasing free radicals. Therefore, physical activity is very important for maintaining the function of the organs. Research has shown that moderate-intensity physical activity on the treadmill reduces liver fibrosis and can repair damaged liver (9). Lindin et al. (2016) stated that moderateintensity aerobic activity reduces the threshold of non-alcoholic hepatic steatosis and liver fibrosis in rats (10). Research has shown that moderate-intensity exercise can repair damaged liver tissue before it is induced by aging (9). In parallel with the increase in the costs of health care in countries, especially in the field of specific groups, the use of non-pharmacological treatments, including therapies and preventive methods of traditional medicine, is expanding rapidly and has become an industry in societies. The most important of these methods is the use of medicinal plants. Citrus aurantium (CA) has great therapeutic potential. These biological effects include anti-cancer. anti-allergic, anti-fat, antibacterial, antioxidant, detoxifying and anti-diabetic effects. The main oils of this plant are limonene, linalool and β -myersen. Photochemically, CA is rich in p-synephrine, alkaloids, and secondary metabolites that affect health. Due to the reduction of physiological processes due to aging and the effect of aging on liver function, physical activity and CA appear to have a beneficial effect on improving metabolic function and delaying the aging process of the liver in the elderly. Therefore, although the cellular mechanisms of exercise and CA activity have not been well identified, the present study

aimed to investigate the effect of aerobic training (T) with CA on SIR1 and PGC-1 α gene expression levels in the liver tissue of elderly rats.

Materials and Methods

In this experimental study, 25 elderly female rats with an average age of 14 months and an average weight of 270- 320 grams were purchased and transferred to the Sports Physiology Laboratory of the Islamic Azad University of Marvdasht Branch. All rats were kept under standard conditions (ith humidity of 45 to 55%, dark-light cycle of 12-12 hours and temperature of 23 \pm 2 ° C) for one week to adapt to the new environment, with free access to water and food. The rats were then divided into five groups of 5 rats, including 1) control, 2) sham, 3) CA, 4) T, and 5) T+CA. Over the course of eight weeks, groups 4 and 5 ran on the treadmill three sessions per week at 65 to 75 percent of the maximum running speed, and groups 3 and 5 received 300 mg/kg/day of CA extract peritoneally. Also, group 2 received 300 mg/kg/day of solvent CA extract (normal saline) peritoneally. At the end of the study, 48 hours after the last training session and CA were anesthetized injection. rats with ketamine and xylazine, and after extraction, the liver tissue was placed in a nitrogen tank and sent to the laboratory to measure SIRT1 and PGC-1 α gene expression levels. SIRT1 and PGC-1 α gene expression levels were measured by real-time PCR. The sequence of SIRT1 and PGC-1 α primers with control gene (TBP) is reported in Table 1. Researchers received introduction letters from Marvdasht Branch of Islamic Azad University with ethics code IR.IAU.M.REC.1399.032.

Aerobic training protocol

At first, the graded exercise testing protocol was taken with a slope of zero degrees to estimate the maximum running speed. To perform this test, the rats initially run at a speed of 10 meters per minute and then their speed increased by 1 meter per minute for every 1 minute. This process continued until the rats were not able to run anymore (exhaustion stage). After determining the maximum running speed, the rats first heated for 5 minutes at 50 to 60 percent of the maximum speed on the treadmill, then they performed continuous training at 65% of the maximum speed in the first week; 70% of the maximum speed in the second week; and 75% of the maximum speed from the third week onwards. In the end, the rats cooled for 5 minutes at a maximum speed of 50 to 60%.

Genes	Primer Sequences	Sizes (Bp)
TBP	Forward: 5'-GCGGGGGTCATGAAATCCAGT -3'	147
	Reverse: 5'-AGTGATGTGGGGGACAAAACGA -3'	
SIRT1	Forward: 5'-TCCTGTGGGATACCTGACTT-3'	300
	Reverse: 5'-AAAGGAACCATGACACTGAATGA -3'	
PGC1a	Forward: 5'-CAGAAGCAGAAAGCAATTGAAGA -3'	230
	Reverse: 5'-GTTTCATTCGACCTGCGTAAAG -3'	

Table 1. The sequence of SIRT1 and PGC-1 α primers with control gene

CA preparation

To prepare the CA extract, 50 g of the powdered sample of the plant was first weighed by a digital scale and added to a balloon containing 500 ml of distilled water connected to a clevenger apparatus. Extraction was performed for 4 hours and the collected essential oil was dehydrated by waterless sodium sulfate, and the prepared essential oil was stored in a freezer at -20° C until use.

Data analysis procedure

The Shapiro-Wilk test was used to evaluate the normal distribution of the findings and one-way ANOVA along with Tukey's *posthoc* test were used to analyze the findings (P <0.05).

Results

The SIRT1 and PGC-1 α gene expression levels are presented in Figures 1 and 2,

respectively. The results of one-way analysis of variance showed a significant difference in SIRT1 and PGC-1 α gene expression levels in the five research groups (P = 0.001).

The results of Tukey's post- hoc test showed that although there was no significant difference in SIRT1 (P = 0.99) and PGC-1 α (P = 0.99) gene expression levels between control and sham groups, SIRT1 gene expression levels in the T group were significantly higher than the control (P =0.009), CA (P = 0.001) and T + CA (P = 0.001) groups. The levels were also significantly lower in the CA group than the control group (p=0.001) (Figure 1). PGC-1a gene expression levels in the CA, T, and T +CA groups were significantly higher than the control group (P = 0.001); in the CA and T + CA groups, the levels significantly higher than the T group (P = 0.001), and in the T + CA group, the levels were significantly higher the than CA group (Figure 2).



Figure 1. SIRT1 gene expression levels in the five groups of study # # # P≤0.001 Significant decrease compared to the control group ** P≤0.01 Significant increase compared to the control group + + + P≤0.001 Significant increase compared to the CA and T+CA groups CA: Citrus aurantium and T: training



Figure 2. PGC-1α gene expression levels in the five groups of study *** P≤0.001 Significant increase compared to the control group + + + P≤0.001 Significant increase compared to the T group \$\$\$ P≤0.001 Significant increase compared to the CA group CA: Citrus aurantium and T: training

Discussion

The results of the present study showed that T significantly increased SIRT1 and PGC-1a gene expression levels in the liver tissue of elderly rats. Aging is associated with a gradual decline in cell biochemical functions such as oxidative phosphorylation and mitochondrial respiration chain. Decreased mitochondrial function leads to increased oxidative stress and leads to aging and modulation at the level of life-enhancing proteins such as SIRT1 (13). In mammals, three members of the sirtoin family (SIRT1, SIRT3, and SIRT6) are involved in aging (14). Excessive expression of SIRT1 in mammals increases life expectancy as well as health in old age (14). Studies show that these epigenetic changes are very dynamic and are influenced by a variety of biological factors, including aging, nutrition, and exercise. In addition, the regulation of SIRT1 by aerobic

exercise regulates PGC-1 α , p53, and NF-_kB well tumor suppressors. as as other transcription factors through diastylase activity (15). Exercise has been shown to reduce oxidative damage to DNA and alter mitochondrial function due to aging by regulating PGC-1 and SIRT1 (16). The results of the present study are consistent with previous studies that have shown that regular exercise increases the expression of SIRT1 and other cytotoxins in many tissues, thereby preventing metabolic diseases or age-related disorders (17). In addition, Van Liu et al. (2019) stated that aerobic exercise increases the expression of SIRT1 in the liver and kidneys of diabetic mice and reduces inflammation and metabolic disorders (18). Another result of the present study was an increase in PGC-1 α expression in liver tissue following T. Ziegler et al. (2019) showed in a study that both aerobic and resistance training increase the expression of PGC-1 α in the

visceral adipose tissue of rats (19). Endurance training using calcium- and phosphatedependent pathways activates adenosine monophosphate and Calmodulin-dependent kinase enzymes, resulting in activation of PGC-1a. Aerobic activity also reduces intracellular energy charge and subsequently activates AMPK and activates intracellular PGC-1 α to increase the expression of genes involved in mitochondrial biogenesis and even increase PGC-1 α (20). However, a study showed that voluntary exercise did not have a significant effect on the protein content of PGC-1 α slow-twitch muscle and liver function (21). Perhaps the difference in the training protocol and the animal model under study has led to differences in results.

In the present study, an increase in SIRT1 expression was associated with increased liver PGC-1a in older rats. Activation of SIRT1 leads to diastylation and activation of PGC- 1α , followed by mitochondrial biogenesis. Increased expression of PGC-1a has been shown to facilitate mitochondrial biogenesis and enhance oxidative phosphorylation in the muscle, heart, and fat tissues (22). The expression of PGC-1 α is also essential to the antioxidant effects of exercise against cell damage caused by aging in the mitochondria and cellular apoptosis (23). Therefore, SIRT1 is an important mediator in the regeneration of cellular metabolism and suppresses inflammatory signals. SIRT1 is also involved controlling hepatic fat metabolism, in modulating hepatic oxidative stress, and protecting against hepatic oxidative stress due to HFD, glucose intolerance, and hepatic steatosis (24). Therefore, treatment strategies to increase SIRT1 levels could potentially protect liver function.

Another result of this study was a significant reduction in liver SIRT1 expression following CA consumption. Flavonones are the main flavonoids in CA. The most abundant free flavonones identified in VA are hesperin and naringin (25). Contrary to the findings of the present study, Shokri Afra et al. (2019) hesperin showed that and resveratrol increased the amount and activity of SIRT1 protein as well as increased phosphorylation of AMPK, although the effect of hesperin was greater than resveratrol (26). Another clinical has shown that hesperin study has antitrogenic effects, regulates fat metabolism, and improves metabolic disorders (27). The effects of CA appear to be mediated by modification in SIRT1. However, the results of the present study showed that CA reduced the expression of SIRT1 in the liver tissue. Perhaps the type of tissue examined or the age of the rats made a difference in the results, which requires further investigation. Studies also show that protein levels and SIRT1 expression are not necessarily the same (28). Therefore, CA may stimulate SIRT1 activity and protein, although it does not affect SIRT1 expression. In the present study, the of liver PGC-1 α increased expression significantly after CA consumption and a combination of aerobic training and CA. The results of a study showed that the glycosidase flavonoid in citrus fruits can activate the AMPK-SIRT1-PGC-1α pathway and thus play a role in controlling fat metabolism (29). Naringin has also been shown to increase PGC-1a, CPT1 and adiponectin expression (30). It appears that the naringin present in CA can increase the expression of PGC-1a and PGC-1 β by increasing thermogenesis by UCP1 (30).

In the present study, the simultaneous effects of T and CA on PGC-1 α were greater than each one alone. Perhaps the two together have synergistic effects and could play a better role in improving the metabolic liver disorder caused by aging. One of the limitations of the present study is the inability to measure SIRT1 and PGC-1 α protein levels by ELISA and Western blot methods, so it is recommended that future studies should examine the effect of T with different intensities and CA consumption with different doses on SIRT1 and PGC-1a protein levels.

Conclusion

Although eight weeks of T and CA consumption alone appear to improve PGC- 1α gene expression levels in the liver tissue of elderly rats, simultaneous T and CA consumption has more favorable effects than each one alone.

References

1. Kim H, Kisseleva T, Brenner DA. Aging and liver disease. Current opinion in gastroenterology 2015;31:184.

[DOI:10.1097/MOG.000000000000176]

2. Okudan N, Belviranli M. Effects of exercise training on hepatic oxidative stress and antioxidant status in aged rats. Archives of physiology and biochemistry 2016;122:180-5. [DOI:10.1080/13813455.2016.1199574]

3. Hunt NJ, Kang SW, Lockwood GP, Le Couteur DG, Cogger VC. Hallmarks of aging in the liver. Computational and Structural Biotechnology Journal 2019. [DOI:10.1016/j.csbj.2019.07.021]

4. Ogrodnik M, Miwa S, Tchkonia T, Tiniakos D, Wilson CL, Lahat A, et al. Cellular senescence drives age-dependent hepatic steatosis. Nature communications 2017;8:1-12. [DOI:10.1038/ncomms15691]

5. Martins R, Lithgow GJ, Link W. Long live FOXO: unraveling the role of FOXO proteins in aging and longevity. Aging cell 2016;15:196-207. [DOI:10.1111/acel.12427]

6. Hunt NJ, McCourt PA, Le Couteur DG, Cogger VC. Novel targets for delaying aging: the

importance of the liver and advances in drug delivery. Advanced drug delivery reviews 2018;135:39-49.

[DOI:10.1016/j.addr.2018.09.006]

7. Jin J, Iakova P, Jiang Y, Medrano EE, Timchenko NA. The reduction of SIRT1 in livers of old mice leads to impaired body homeostasis and to inhibition of liver proliferation. Hepatology 2011;54:989-98. [DOI:10.1002/hep.24471]

8. Haigis MC, Sinclair DA. Mammalian sirtuins: biological insights and disease relevance. Annual Review of Pathology: Mechanisms of Disease 2010;5:253-95.

[DOI:10.1146/annurev.pathol.4.110807.092250]

9. Wasityastuti W, Habib NA, Sari DC, Arfian N. Effects of low and moderate treadmill exercise on liver of d-galactose-exposed aging rat model. Physiological reports 2019;7:e14279. [DOI:10.14814/phy2.14279]

10. Linden MA, Sheldon RD, Meers GM, Ortinau LC, Morris EM, Vieira-Potter VJ, et al. Exercise Training As A Mitigator Of Liver Fibrosis In Western Diet Fed OLETF Rats: 1783 Board# 3 June 2, 1: 00 PM-3: 00 PM. Medicine and science in sports and exercise 2016;48:485-. [DOI:10.1249/01.mss.0000486456.42848.e4]

11. Yazdanparast Chaharmahali B, Azarbayjani MA, Peeri M, Farzanegi Arkhazloo P. The Effect of Moderate and High Intensity Interval Trainings on Cardiac Apoptosis in the Old Female Rats. Report of Health Care 2018;4:26-35.

12. He W, Li Y, Liu M, Yu H, Chen Q, Chen Y, et al. Citrus aurantium L. and its flavonoids regulate TNBS-induced inflammatory bowel disease through anti-inflammation and suppressing isolated jejunum contraction. International journal of molecular sciences 2018;19:3057. [DOI:10.3390/ijms19103057]

13. Tonkin J, Villarroya F, Puri PL, Vinciguerra M. SIRT1 signaling as potential modulator of skeletal muscle diseases. Current opinion in pharmacology 2012;12:372-6. [DOI:10.1016/j.coph.2012.02.010] 14. Herranz D, Muñoz-Martin M, Cañamero M, Mulero F, Martinez-Pastor B, Fernandez-Capetillo O, et al. Sirt1 improves healthy ageing and protects from metabolic syndrome-associated cancer. Nature communications 2010;1:1-8. [DOI:10.1038/ncomms1001]

 Ntanasis-Stathopoulos J, Tzanninis J, Philippou A, Koutsilieris M. Epigenetic regulation on gene expression induced by physical exercise.
J Musculoskelet Neuronal Interact 2013;13:133-46.

16. Fiuza-Luces C, Garatachea N, Berger NA, Lucia A. Exercise is the real polypill. Physiology 2013;28:330-58.

[DOI:10.1152/physiol.00019.2013]

17. Suwa M, Sakuma K. The potential role of sirtuins regarding the effects of exercise on aging-related diseases. Current aging science 2013;6:178-88. [DOI:10.2174/18746098112059990035]

18. Liu H-W, Kao H-H, Wu C-H. Exercise training upregulates SIRT1 to attenuate inflammation and metabolic dysfunction in kidney and liver of diabetic db/db mice. Nutrition & metabolism 2019;16:22. [DOI:10.1186/s12986-019-0349-4]

19. Ziegler A, Damgaard A, Mackey A, Schjerling P, Magnusson P, Olesen A, et al. An anti-inflammatory phenotype in visceral adipose tissue of old lean mice, augmented by exercise. Scientific reports 2019;9:1-10. [DOI:10.1038/s41598-019-48587-2]

20. Roberts-Wilson TK, Reddy RN, Bailey JL, Zheng B, Ordas R, Gooch JL, et al. Calcineurin signaling and PGC-1 α expression are suppressed during muscle atrophy due to diabetes. Biochimica et Biophysica Acta (BBA)-Molecular Cell Research 2010;1803:960-7. [DOI:10.1016/j.bbamcr.2010.03.019]

21. Matiello R, Fukui RT, Silva ME, Rocha DM, Wajchenberg BL, Azhar S, et al. Differential regulation of PGC-1 α expression in rat liver and skeletal muscle in response to voluntary running.

Nutrition & metabolism 2010;7:36. [DOI:10.1186/1743-7075-7-36]

22. Arany Z, He H, Lin J, Hoyer K, Handschin C, Toka O, et al. Transcriptional coactivator PGC-1 α controls the energy state and contractile function of cardiac muscle. Cell metabolism 2005;1:259-71. [DOI:10.1016/j.cmet.2005.03.002]

23. Leick L, Lyngby SS, Wojtasewski JF, Pilegaard H. PGC-1 α is required for traininginduced prevention of age-associated decline in mitochondrial enzymes in mouse skeletal muscle. Experimental gerontology 2010;45:336-42. [DOI:10.1016/j.exger.2010.01.011]

24. Ding R-B, Bao J, Deng C-X. Emerging roles of SIRT1 in fatty liver diseases. International journal of biological sciences 2017;13:852. [DOI:10.7150/ijbs.19370]

25. Lee SH, Yumnam S, Hong GE, Raha S, Venkatarame Gowda Saralamma V, Lee HJ, et al. Flavonoids of Korean Citrus aurantium L. induce apoptosis via intrinsic pathway in human hepatoblastoma HepG2 cells. Phytotherapy research 2015;29:1940-9. [DOI:10.1002/ptr.5488]

26. Afra HS, Zangooei M, Meshkani R, Ghahremani MH, Ilbeigi D, Khedri A, et al. Hesperetin is a potent bioactivator that activates SIRT1-AMPK signaling pathway in HepG2 cells. Journal of physiology and biochemistry 2019;75:125-33. [DOI:10.1007/s13105-019-00678-4]

27. Jayaraman R, Subramani S, Abdullah SHS, Udaiyar M. Antihyperglycemic effect of hesperetin, а citrus flavonoid, extenuates hyperglycemia and exploring the potential role in antihyperlipidemic antioxidant and in streptozotocin-induced diabetic rats. Biomedicine & Pharmacotherapy 2018;97:98-106. [DOI:10.1016/j.biopha.2017.10.102]

28. Sasaki T, Maier B, Koclega KD, Chruszcz M, Gluba W, Stukenberg PT, et al. Phosphorylation regulates SIRT1 function. PloS one 2008;3. [DOI:10.1371/journal.pone.0004020] 29. Wu H, Liu Y, Chen X, Zhu D, Ma J, Yan Y, et al. Neohesperidin exerts lipid-regulating effects in vitro and in vivo via fibroblast growth factor 21 and AMP-activated protein Kinase/Sirtuin Type 1/Peroxisome Proliferator-activated receptor gamma coactivator 1α signaling axis. Pharmacology 2017;100:115-26. [DOI:10.1159/000452492]

30. Rebello CJ, Greenway FL, Lau FH, Lin Y, Stephens JM, Johnson WD, et al. Naringenin promotes thermogenic gene expression in human

white adipose tissue. Obesity 2019;27:103-11. [DOI:10.1002/oby.22352]

How to cite:

Shykholeslami Z, Abdi A, Barari A, Hosseini S.A. The effect of aerobic training with Citrus aurantium L. on Sirtuin 1 and peroxisome proliferator-activated receptor gamma coactivator 1-alpha gene expression levels in the liver tissue of elderly rats. Jorjani Biomedicine Journal. 2020; 8(1): 57-65.