

Online ISSN: 2645-3509

In-vitro antibacterial effects of Artemisia extracts on clinical strains of *P. aeruginosa*, *S. pyogenes*, and *oral bacteria*

Fatemeh Monirian¹, Reyhane Abedi¹, Negar Balmeh^{2*}, Samira Mahmoudi², Fereshteh Mirzaei Poor²

1. Farzanegan 2 Nezhad Satari high school, Isfahan, Iran

2. Shahriari Research Center, Isfahan, Iran

Article Type: Original Article

Article History:

Received: 24 Jul 2020 Revised: 1 Aug 2020 Accepted: 4 Aug 2020

*Correspondence:

Negar Balmeh, Shahriari Research Center, Isfahan, Iran.

n.balmeh@gmail.com



DOI: 10.29252/jorjanibiomedj.8.3.35

Abstract

Background and objective: Some common problems in the health care system are Microbial resistance to antibiotics, the side effects of food additives, and preservatives. Considering the antibiotic resistance of microorganisms and the need to identify new compounds, the present study was conducted to determine the antimicrobial effects of *Artemisia* extracts.

Material And Method The study was performed in two stages including extraction and determination of antibacterial properties of aqueous, ethanolic, methanolic, acetone/ethanolic, and hydroethanolic extracts of *Artemisia* on standard *Pseudomonas aeruginosa*, *Streptococcus pyogenes*, and oral bacteria sample. The well diffusion method was performed to evaluate the concentration that had an inhibitory and bactericidal effect.

Result: Aqueous extract had an effect on the oral bacteria sample (8 mm), the methanolic extract had an effect on *S. pyogenes* and oral bacteria sample (15 mm) and finally acetone/ethanolic extracts had antimicrobial properties against *S. pyogenes* (16 mm). The concentration used for all five extracts was 50 mg/ml and showed an inhibition effect on the growth of *S. pyogenes* standard strain and oral bacteria sample. The less serial dilutions of extracts were tested but no antibacterial effects were seen. So, 50 mg/ml was the minimum concentration that had an inhibitory and bactericidal effect.

Conclusion: It can be inferred that aqueous, methanolic, and acetone/ethanolic extracts of *Artemisia* had the highest inhibitory effect on *S. pyogenes* and the oral bacteria sample. Consequently, by applying different extraction methods and by utilizing different solvents, it may be possible to more efficiently obtain biomaterials with antimicrobial properties from this plant.

Keywords: herbal antibacterial effects, *Artemisia extracts, Pseudomonas aeruginosa, Streptococcus pyogenes,* oral bacteria.

Copyright© 2018, **Jorjani Biomedicine Journal** has published this work as an open access article under the terms of the Creative Commons Attribution License (<u>http://creativecommons.org/licenses/by-nc/4.0/</u>) which permits non-commercial uses of the work while it is properly cited.



Introduction

In the new century, the tendency of consumers to use fewer chemicals and preservatives has led to extensive research on the benefits and harms of using natural compounds with antimicrobial properties. Further, the resistance of microorganisms to antibiotics poses problems that have become one of the major concerns of human society and the health care system (1). With the advent of antibiotic-resistant bacteria, the effectiveness of existing drugs has diminished, increasing the failure of antimicrobial treatments. In addition, the widespread use of industrial drugs, as well as the misuse of these drugs, can lead to many side effects (2,3). As a result, one way to prevent the development of pathogenic bacteria resistant to antibiotics is to use new compounds that are fundamentally different from the existing industrial source in terms of basic structure with antimicrobial drugs (4). Plants produce secondary metabolites flavonoids, alkaloids, including tannins. glycosides, terpenes, and phenolic compounds. Some of these compounds have molecular complex structures and antimicrobial properties, therefore, in recent years, there has been a strong desire to discover and introduce antimicrobial agents of plant origin (5). Knowledge of chemical compounds in plants is important not only for the discovery of medicinal drugs but also because the use of chemical products of plant origin that have antimicrobial properties help control resistant strains (6). In this regard, the World Health Organization has recommended the use of natural drugs as a reliable method for the treatment of microbial and nonmicrobial diseases (7). Numerous plants have been identified that have antimicrobial properties, but most of them have not yet been adequately studied (8). Plants as a source of new drugs remain largely unknown, and screening studies to discover the effects of their biologically active compounds and molecules are necessary. The first step in achieving this goal is to investigate their antimicrobial activity in vitro (9).

Artemisia is one of the medicinal plants that is used in traditional medicine to treat some diseases and so far many studies have been performed on it to detect different degrees of inhibitory and antimicrobial effects. The plant is widely grown in a variety of climates, especially in arid regions where plant growth somehow difficult climatic due to is conditions; one of the reasons for this expansion could be the high ability of this plant to adapt to different climatic conditions. Chemically, Artemisia extracts often contain polypranoids and aromatic compounds that can have antimicrobial effects on a range of bacteria due to their phenolic groups in their structure (10).

Although the determination of the active ingredients of the plants is pharmacologically important, it is necessary to first determine the effectiveness of the extracts obtained from them (11). In this regard, it has been noted that in the early stages of the discovery and production of new drugs, the selection of raw plant extracts may be more appropriate than the screening of compounds for this reason (12). In the present study, we conducted a preliminary screening to determine the antimicrobial effect of aqueous, ethanolic, methanolic. acetone/ethanolic and hydroethanolic extracts of Artemisia aqueous on standard P. aeruginosa, S. pyogenes, and oral bacteria sample from rotten teeth, in order to investigate the antibacterial effect of Artemisia for using as a natural mouth wash or antibacterial agent as preservative material in food, cosmetic, and the health industries.

Materials and Methods

Preparing the plant

Artemisia specimens were collected from the Isfahan Medicinal plants market. After collection, the sample was rinsed several times with water, then dried in the shade, and the sample was powdered with a grinder for easy extraction.

Extraction

The extract of the plant prepared in the separate stages with solvents including water, ethanol, hydroethanol, acetone/ethanol, and methanol. Extraction was done by the maceration method for all solvents, but hydroethanolic extract was performed using the Soxhlet method to determine whether the temperature in the extraction process affects the antibacterial properties of the extracts. To prepare the water extract, 24 grams of plant powder was added into an Erlenmeyer flask and increased the volume with distilled water to 100 ml. It was put in a shaker for 3 days, then the extract was first filtered with a sterile lace cloth and then with filter paper. After smoothing, the solution was poured into a petri dish and placed in an incubator at 37 C to evaporate water and dry completely (13). After drying, the extract was kept away from light. The above method was also used to ethanolic. methanolic. prepare and acetone/ethanolic extracts. For preparing hydroethanolic extracts, 24 g of dried Artemisia powder with 200 ml hydroethanolic solvent was placed in the Soxhlet extractor for 7 hours. The solvent was evaporated slowly and was concentrated at 37 C. Then, 5% of Dimethyl sulfoxide (DMSO) (Sigma-Aldrich, United States) was utilized to prepare the extract. Finally, two types of solvents including water and DMSO were used to dissolve the various extracts. The aqueous extract of Artemisia in water and other extracts in DMSO were completely dissolved and finally filtered with 0.22 μ m MS ® MCE Syringe Filter from Membrane Solutions Company (14,15).

Evaluation of the antibacterial activity of the extract

In the present study, P. aeruginosa and S. pyogenes, and the oral bacteria sample were used to evaluate the antibacterial activity of Artemisia extract. The oral bacterial sample was randomly chosen by a general sampling of a decayed tooth and no specific strains identification was done. P. aeruginosa and S. pyogenes were clinical strains and were provided from the Al-Zahra University Hospital of Isfahan, Iran. First, equal volumes of each bacterial suspension were prepared according to standard 0.5 McFarland (16) vy7`. The bacteria were first cultured on Mueller Hinton Agar medium (QUELAB's, Canada). To investigate the antibacterial activity of each extract on studied bacteria, extracts were used at a concentration of 50 mg/ml. Dilution antimicrobial experiments were performed twice and in accordance with the recommendations of the Standards Committee of Clinical Laboratories (CLSI) (17). For this purpose, 15 ml of sterile culture medium with an appropriate temperature was transferred into 8 mm Petri dishes. The surface of the culture medium was covered with bacterial suspension, then 5 well with a diameter of 6 mm were made in each plate using a sterile Pasteur pipette. To assure the accuracy of the test and comparison conditions, description gentamicin and nalidixic acid antibiotic discs (Padtan Teb Company, Iran) were settled in two wells (positive control). DMSO was also utilized as a negative control sample in one of the wells. The plant extract suspension was loaded into two wells. Each extract was analyzed on P. aeruginosa, S. pyogenes, and the oral bacteria sample. Finally, all the plates were incubated at 37 °C for 24 hours.

After finding the Artemisia extracts that had antibacterial effects. the different concentrations of those extracts were prepared and poured in the prepared wells on Mueller Hinton Agar medium. Seven vials were prepared. 900 microliters of distilled water was poured into each vial. About 100 µl was obtained from the main vial with a concentration of 50 mg/ml and added to 900 ul of distilled water in the first vial and well mixed. Next, 100 µl were removed from the first vial and added to the second vial, and this process continued until the seventh vial. Finally, 100 µl was removed from the last vial and poured out. In this way, a series of dilutions from the 50 mg/ml of extracts were prepared. An equal amount of the extract in each vial was transferred to wells in Mueller Hinton Agar medium containing the cultured bacteria to determine the antibacterial properties of each concentration. All plates incubated at 37 ° C for 24 h. Then the lowest concentration that caused the non-growth zone was considered as the minimum inhibitory dosage. Units from the non-growth area of each dilution were collected and to ACC at the position 1003. In fact, nucleotide transitional mutation of G to A has caused alanine 335 amino acid to be replaced by threonine.

Result

The gained results have revealed that aqueous extract of *Artemis*ia had no effects on *P*. *aeruginosa* and *S. pyogenes* but it has an inhibiting effect on oral bacteria sample. The ethanolic and hydroethanolic extract of *Artemisia* did not indicate any anti-bacterial effects on all three mentioned bacteria strains. The result of the methanolic extract has shown more impact on different bacteria.

As demonstrated in <u>Table 1</u>, the methanolic extract had a significant antibacterial effect on *S. pyogenes* and the oral bacteria sample. Also, the acetone/ ethanolic extract had the most inhibiting effect on *S. pyogenes*.

50 mg/ml of *Artemisia* extracts was the minimum concentration, which had inhibitory and bactericidal effects. None of the fewer concentrations had an antibacterial effect. (Table 2)

Concentration (mg/ml) Bacteria Strain	Aqueous extract (Maceration) 50 mg/ml	Ethanolic extract (Maceration) 50 mg/ml	Methanolic extract (Maceration) 50 mg/ml	Acetone/ ethanolic extract (Maceration) 50 mg/ml	Hydro ethanolic extract (Soxhlet) 50 mg/ml	Nalidixic acid	Gentami cin
P. aeruginosa	0	0	0	0	0	0	25
S. pyogenes	0	0	15 ± 0.21	16 ± 0.09	0	15 ± 0.15	30 ± 0.14
oral bacteria sample	8 ± 0.1	0	15 ± 0.37	0	0	N/D	N/D

Table 1. Mean diameter of growth inhibitory zone of aqueous, ethanolic, methanolic, acetone/ ethanolic, and hydroethanolic extracts of *Artemisia* against selected bacteria in millimeters (mean ± standard deviation).

Table 2. The minimum concentration of aqueous, ethanolic, methanolic, acetone/ethanolic, and hydroethanolic extracts of *Artemisia* (mg/ml) with bacterial growth inhibitory and bactericidal effect.

Substance Bacteria Strain	Aqueous extract	Ethanolic extract	Methanolic extract	Acetone/ethanolic extract	Hydroethanolic extract
P. aeruginosa	-	-	-	-	-
S. pyogenes	-	-	50	50	-
oral bacteria sample	50	-	50	-	-

Discussion

Artemisia extract has been reported to contain many antimicrobial compounds such as phenols, cineol, ketones, camphor, and thymol, all of which interfere with the action of membrane enzymes by destroying cell walls and bacterial proteins; in fact, the presence of antibacterial activity of this plant can be attributed to the presence of these compounds (10,19,20). In Artemis species, there are high concentrations of beneficial substances along with antioxidants that are effective in maintaining the natural flora of the body. These antimicrobial effects are likely to be mediated by a variety of mechanisms, including disruption of the mitochondrial-related proton chain, disruption of membrane proteins and lipids, and inhibition of the sarcoplasmic reticulum calcium pump (21).

According to the findings of a survey, the quality of natural extracts and their antioxidant effects depend not only on storage time, geographical origin, and harvest time but also on the environment and technological factors used for the extraction process. Also, the type of solvent used in extraction is one of the other important factors that are important in causing antioxidant effects (22). The results obtained by Appalasamy *et al.* Also showed that the extract obtained from *Artemisia* at a concentration of 6 mg/ml led to inhibitory

effects on Escherichia coli and Staphylococcus. This result proves that the active ingredients in the crude extract have synergistic and intensifying effects, thus resulting in a higher overall antimicrobial activity (23). Another study by Jutea et al. revealed that Artemis extract significantly inhibited the growth of Enterococcus hirae (24). Findings of a study also stated that various extracts prepared from Artemis leaves have significant antioxidant properties compared to the standard curve of vitamin C and ferrous sulfate (25).

In a study conducted on Artemisia from Kerman province of Iran, it was found that the essential oil of this plant has antibacterial and Minimum fungal effects. inhibitory concentrations were obtained for Staphylococcus aureus (4 µg/ml), Salmonella typhi (32 µg/ml), E. coli (32 µg/ml), Candida albicans (8 µg/ml), and Aspergillus niger (4 µg/ml) (26). Another study on Artemisia from the city of Babol in Iran also confirmed that aqueous, ethanolic, and methanolic all extracts of Artemisia were able to have significant antibacterial properties in comparison with the blank and antibiotic sample. Aqueous extract with a 30% antibacterial effect of ciprofloxacin on E. coli and methanolic extract with the highest antimicrobial activity with 71% of vancomycin showed an inhibitory effect on the growth Enterococcus faecalis. of

Although the aqueous extract had the least inhibitory effect on E. coli and Enterococcus faecalis, the inhibitory effects of this extract were significant compared to the blank sample (27). Based on an in vitro study on aqueous, ethanolic, and methanolic of Artemisia from Markazi province of Iran, it was identified that their MIC were 6250, 3125, and 6250 µg/ml, respectively, so it was concluded that different extracts affect Candida albicans growth and prevent its growth (28). The results of an investigation on Artemisia From the western regions of Isfahan province, Iran, determined that flower essential oil had an inhibitory effect on the growth of pathogenic bacteria, especially gram-positive types (Staphylococcus aureus, Enterococcus faecalis, E. coli) because the diameter of the non-growth zone for was in the range of 8.3-45 mm and the least MIC was 2.5%. However, this essential oil did not affect P. aeruginosa (29).

In the present study, the bacterial effects of different concentrations of aqueous, ethanolic, methanolic, acetone/ethanolic, and hydroethanolic extracts of *Artemisia* under standard laboratory conditions on *P. aeruginosa*, *S. pyogenes*, and oral bacteria sample were investigated.

In most Iranian previous studies, one or two types of solvents have been used for extraction, while in the present study, attempts were made to use different types of *Artemisia* extracts for investigation. Also, to select the bacterial strain, the species that had not been used in Iranian studies on *Artemisia* were selected.

Taking into account these two factors (the simultaneous study of several types of extracts and selection of untested strains), it was tried to obtain new and complete information about the antibacterial effect of this plant and to use it for pharmaceutical studies.

The findings showed that aqueous, methanolic, and acetone/ethanolic extracts of Artemisia had significant antibacterial effects on S. pyogenes. All effective extracts at a concentration of 50 mg had antimicrobial effects. Therefore. by using different extraction methods and various solvents, it may be possible to more effectively obtain biomaterials with antimicrobial properties from this plant. Considering that both in this study and in one of the previous studies on the essential oil of the Iranian Artemisia plant, it was found that they have less antimicrobial effects on P. aeruginosa, it seems that in future studies, higher concentrations of these extracts are better to be used for assessment. But overall, it can be concluded with certainty that this plant has a significant effect on gram-positive bacteria and is, therefore, a good choice for use in the food and pharmaceutical industries.

The oral bacterial sample was also randomly selected by a general sampling of a decayed tooth and no specific identification of the strains was performed to evaluate *Artemisia* antimicrobial overall effect on tooth decaying bacteria. And because of the positive results obtained from inhibitory tests, *Artemisia* extracts may be used in general for producing all types of mouthwashes and toothpaste. It is a better option compared to the chemical compounds used in these products.

Conclusion

It can be deduced that aqueous, methanolic, and acetone/ethanolic extracts of *Artemisia* have antibacterial effects on both *S. pyogenes* bacteria and the oral bacteria sample. This study also revealed that none of the extracts of *Artemisia* had an antibacterial effect on *P*. *aeruginosa*. According to the findings of this study, Artemisia plant extracts can be used in food, cosmetics, and pharmaceutical industries due to its antibacterial properties.

References

1. Stappen I, Wanner J, Tabanca N, Wedge DE, Ali A, Khan IA, et al. Chemical composition and biological effects of Artemisia maritima and Artemisia nilagirica essential oils from wild plants of western Himalaya. Planta Med. 2014;80(13):1079-87. [DOI:10.1055/s-0034-1382957]

2. Ahameethunisa AR, Hopper W. In vitro antimicrobial activity on clinical microbial strains and antioxidant properties of Artemisia parviflora. Ann Clin Microbiol Antimicrob. 2012;11(1):30. [DOI:10.1186/1476-0711-11-30]

3. Mahmoudi S, Nasiri R, Jafari Sales A. Invitro antibacterial effects of methanolic extract of peppermint (Mentha Piperita Lamiaceae) on standard Staphylococcus aureus, Bacillus cereus, Escherichia coli and Pseudomonas aeruginosa strain. Jorjani Biomed J. :1-7. [DOI:10.29252/jorjanibiomedj.7.4.4]

4. Uniyal SK, Singh KN, Jamwal P, Lal B. Traditional use of medicinal plants among the tribal communities of Chhota Bhangal, Western Himalaya. J Ethnobiol Ethnomed. 2006;2(1):14. [DOI:10.1186/1746-4269-2-14]

5. Arokiyaraj S, Sripriya N, Bhagya R, Radhika B, Prameela L, Udayaprakash NK. Phytochemical screening, antibacterial and free radical scavenging effects of Artemisia nilagirica, Mimosa pudica and Clerodendrum siphonanthus-An in-vitro study. Asian Pac J Trop Biomed. 2012;2(2):S601-4. [DOI:10.1016/S2221-1691(12)60281-0] 6. Salehi S, Mirzaie A, Sadat Shandiz SA, Noorbazargan H, Rahimi A, Yarmohammadi S, et al. Chemical composition, antioxidant, antibacterial and cytotoxic effects of Artemisia marschalliana Sprengel extract. Nat Prod Res. 2017;31(4):469-72. [DOI:10.1080/14786419.2016.1174234]

7. Kalaivani R, Devi VJ, Umarani R, Periyanayagam K, Kumaraguru AK. Antimicrobial Activity of Some Important Medicinal Plant oils against Human Pathogens. J Biol Act Prod from Nat. 2012;2(1):30-7.

DOI:10.1080/22311866.2012.10719105

8. Fallah N, Namazi S, Balmeh N, Mahmoudi S, Mirzaei Poor F. In-vitro Antibacterial Effects of Aqueous, Ethanolic, Methanolic, Acetone and Hydro Ethanolic Extracts of Achillea Millefolium on Standard Klebsiella, S. pyogenes, and Oral Bacterias Strains. Tabari Biomed Student Res J [Internet]. 2020;2(3):22-7. Available from: http://tbsrj.mazums.ac.ir/article-1-3711-en.html [DOI:10.18502/tbsrj.v2i3.4529]

9. Rawani A, Pal S, Chandra G. Evaluation of antimicrobial properties of four plant extracts against human pathogens. Asian Pac J Trop Biomed. 2011;1(1):S71-5. [DOI:10.1016/S2221-1691(11)60127-5]

10. Habibi Z, Ghanian S, Ghasemi S, Yousefi M. Chemical composition and antibacterial activity of the volatile oil from seeds of Artemisia annua L. from Iran. Nat Prod Res. 2013;27(2):198-200.

[DOI:10.1080/14786419.2012.662652]

11. Djeussi DE, Noumedem JAK, Seukep JA, Fankam AG, Voukeng IK, Tankeo SB, et al. Antibacterial activities of selected edible plants extracts against multidrug-resistant Gram-negative bacteria. BMC Complement Altern Med. 2013;13(1):164. [DOI:10.1186/1472-6882-13-164] 12. Rojas JJ, Ochoa VJ, Ocampo SA, Muñoz JF. Screening for antimicrobial activity of ten medicinal plants used in Colombian folkloric medicine: A possible alternative in the treatment of non-nosocomial infections. BMC Complement Altern Med. 2006;6(1):1-6. [DOI:10.1186/1472-6882-6-2]

13. Mashhadian N V, Rakhshandeh H. Antibacterial and antifungal effects of Nigella sativa extracts against S. aureus, P. aeroginosa and C. albicans. Pak J Med Sci. 2005;21(1):47-52.

14. Grare M, Fontanay S, Cornil C, Finance C, Duval RE. Tetrazolium salts for MIC determination in microplates: Why? Which salt to select? How? J Microbiol Methods. 2008;75(1):156-9.

[DOI:10.1016/j.mimet.2008.05.010]

15. Nazri NAAM, Ahmat N, Adnan A, Mohamad SAS, Ruzaina SAS. In vitro antibacterial and radical scavenging activities of Malaysian table salad. African J Biotechnol. 2011;10(30):5728-35.

16. Islam MA, Alam MM, Choudhury ME, Kobayashi N, Ahmed MU. Determination of minimum inhibitory concentration (MIC) of cloxacillin for selected isolates of methicillinresistant Staphylococcus aureus (MRSA) with their antibiogram. Bangladesh J Vet Med. 2008;6(1):121-6.

[DOI:10.3329/bjvm.v6i1.1350]

17. Osato MS. Antimicrobial susceptibility testing for Helicobacter pylori: sensitivity test results and their clinical relevance. Curr Pharm Des. 2000;6(15):1545-55. [DOI:10.2174/1381612003399059]

18. Kermanshah H, Kamangar H, Arami S, Mirsalehian A, Kamalinegad M, Karimi M, et al. Comparison of antibacterial effect of hydroalcoholic extract of four plants against cariogenic microorganisms by two in vitro Methods. J Babol Univ Med Sci. 2011;13(6):21-9.

19. Gupta PC, Dutta B, Pant D, Joshi P, Lohar DR. In vitro antibacterial activity of Artemisia annua Linn. growing in India. Int J Green Pharm. 2009;3(3). [DOI:10.4103/0973-8258.56287]

20. Friedman M, Henika PR, Mandrell RE. Bactericidal activities of plant essential oils and some of their isolated constituents against Campylobacter jejuni, Escherichia coli, Listeria monocytogenes, and Salmonella enterica. J Food Prot. 2002;65(10):1545-60. [DOI:10.4315/0362-028X-65.10.1545]

21. Brisibe EA, Umoren UE, Owai PU, Brisibe F. Dietary inclusion of dried Artemisia annua leaves for management of coccidiosis and growth enhancement in chickens. African J Biotechnol. 2008;7(22).

22. Moure A, Franco D, Sineiro J, Domínguez H, Núñez MJ, Lema JM. Antioxidant activity of extracts from Gevuina avellana and Rosa rubiginosa defatted seeds. Food Res Int. 2001;34(2-3):103-9. [DOI:10.1016/S0963-9969(00)00136-8]

23. Appalasamy S, Lo KY, Ch'ng SJ, Nornadia K, Othman AS, Chan L-K. Antimicrobial activity of artemisinin and precursor derived from in vitro plantlets of Artemisia annua L. Biomed Res Int. 2014;2014. [DOI:10.1155/2014/215872]

24. Juteau F, Masotti V, Bessiere JM, Dherbomez M, Viano J. Antibacterial and antioxidant activities of Artemisia annua essential oil. Fitoterapia. 2002;73(6):532-5. [DOI:10.1016/S0367-326X(02)00175-2]

25. Baradaran M, Ashrafpour M, Rezaei H, Sefidgar AA, Sharifi H. Antioxidant activity of different extracts of the Artemisia Annua growing in an area of Babol city. 2014; 26. Ghanjali A, Pourramezani Hm. Survey of Antibacterial and Antifungal Impact of Essential Oil of Artemisia Kermanensis.2013;

27. Ashrafpour M, Baradaran M, Sharifi H. Survey of the Antibacterial Properties of Aqueous Ethanolic and Methanolic Extraction of Artemisia Annua Around the City of Babol. Sci J ilam Univ Med Sci. 2016;23(6):129-41.

28. Taheri AA, Gholampourazizi I, Hashemi Karoui M, Farhadi L, Servatyari K, Rouhi S. Inhibitory effect of aquatic and alcoholic extracts of Artemisia sieberi on growth of Candida albicans: an in vitro study. Qom Univ Med Sci J. 2018;12(6):39-47. [DOI:10.29252/qums.12.6.39]

29. Naghsh A, Mohammadi Sichani M, Amjad L. Antibacterial Activity of the Essential Oil of Artemisia deserti Flowers Against Some Pathogenic Bacteria. Qom Univ Med Sci J. 2015;8(5):57-64.

How to cite:

Monirian F, Abedi R, Balmeh N, Mahmoudi S, Mirzaei Poor F. In-vitro antibacterial effects of Artemisia extracts on clinical strains of *P. aeruginosa, S. pyogenes*, and *oral bacteria*. Jorjani Biomedicine Journal. 2020; 8(3): 35-43.